

Technical Data Sheet

Mouse Th1/Th17 Phenotyping Kit

Product Information

Material Number: 560759
Size: 50 Tests

Description

Components:

51-9006632	Mouse Th1/Th17 Phenotyping Cocktail	1.0 ml
	Containing the following: Mouse CD4 PerCP-Cy5.5 (clone: RM4-5) Mouse IL-17A PE (clone: TC11-18H10.1) Mouse IFN-GMA FITC (clone: XMG1.2)	
51-9006613	BD Cytotfix™ Fixation Buffer	100 ml
51-2091KE	BD Perm/Wash™ Buffer	25 ml
51-2092KZ	BD GolgiStop™ Protein Transport Inhibitor (containing monensin)	0.7 ml

The peripheral CD4⁺ T cell pool includes multiple effector and memory T cell subsets that arise through antigen-driven expansion and differentiation of naïve T cells. The early response of naïve CD4⁺ T cells to antigenic stimulation is characterized by high level proliferation and a limited cytokine repertoire. Further differentiation yields cells with a more diverse potential for cytokine expression. Depending upon the balance of local cytokines, costimulatory molecules, antigen levels, and genetic factors, Type-1 T helper (Th1), Th2, and Th17 effector and/or memory cells are generated by immune responses.

Functionally-polarized CD4⁺ T cell subsets have been identified based on their distinctive patterns of cytokine secretion. As a signature cytokine, Th1 cells selectively produce large amounts of interferon-gamma (IFN- γ). Th2 cells selectively produce IL-4, and Th17 express high levels of IL-17A. Through secretion of IFN- γ and other effector molecules, Th1 cells activate macrophages, natural killer (NK) cells, and CD8⁺ T cells and are responsible for cell-mediated immunity. Th1 cells provide protection against intracellular bacteria, fungi, protozoa and viruses and are involved in some autoimmune responses. IL-4 produced by Th2 cells is particularly strong in driving B cells to generate IgE-secreting cells. IgE plays a role in basophil/mast cell mediated immune reactions. Th2 cells mediate protection against extracellular parasites but may also cause harmful allergic responsiveness to develop. Through the secretion of IL-17A and other factors, Th17 cells recruit and activate neutrophils and mediate immune responses against extracellular bacteria and fungi. Th17 cells are also implicated in autoimmune responses. In addition to these types of T helper cells, Th0- (IL-4 and IFN- γ) and Th17/Th1-like (IL-17A/IFN- γ) cells that coexpress signature cytokines have been described.

The Th1/Th17 paradigm provides a useful model system for investigating the cellular and molecular mechanisms that mediate protective as well as harmful immune responses. The BD Mouse Th1/Th17 Phenotyping Kit provides an easy-to-use four-color cocktail of fluorescent antibodies-specific for mouse CD4, IFN- γ (for Th1), IL-4 (for Th2) and IL-17A (for Th17)-that will enable researchers to identify and characterize the nature of these T helper cell types by multicolor flow cytometric analysis. The kit can be used to successfully analyze ex vivo lymphoid cell samples (eg, for the types of in vivo-generated T helper cells) or to monitor T helper cell differentiation by cells cultured within various experimental model systems.

Investigators should note that the appearance of BD GolgiStop™ Protein Transport Inhibitor may range in color from clear (colorless) to light yellow.

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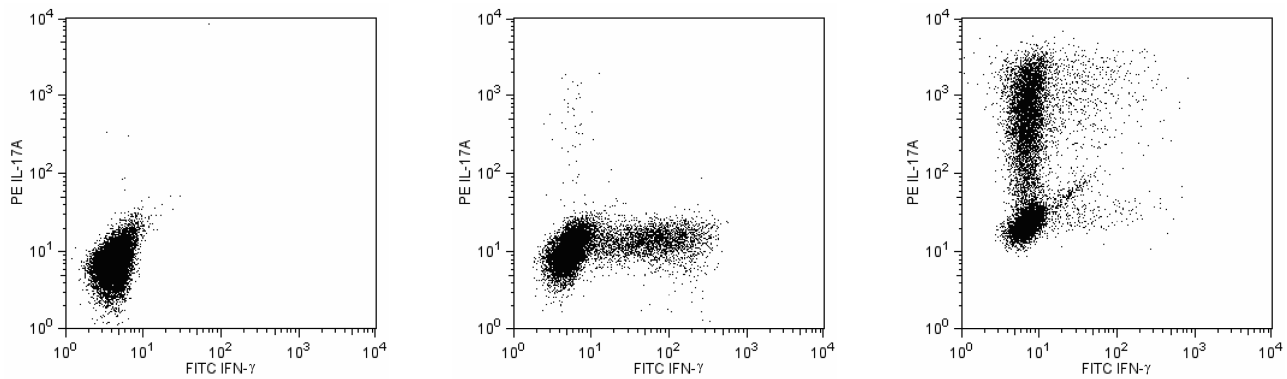
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Flow cytometric analysis of Mouse Th1/Th17 phenotyping kit. The staining pattern of IFN- γ and IL-17A on resting spleen cells (left panel), PMA/Ionomycin stimulated spleen cells (middle panel) and polarized Th17 cells (right panel) are shown. Th17 cells were generated from mouse spleen cells for a 14 days period under polarization condition described by Chen Dong et al (see reference below). Dead cells appear on the diagonal. Dot plot analysis are derived from gated CD4⁺ cells population. Flow cytometry was performed on a BD FACSCalibur™.

Preparation and Storage

Store undiluted at 4°C and protected from prolonged exposure to light. Do not freeze.

The monoclonal antibody was purified from tissue culture supernatant or ascites by affinity chromatography.

The antibody was conjugated with FITC under optimum conditions, and unreacted FITC was removed.

The antibody was conjugated with R-PE under optimum conditions, and unconjugated antibody and free PE were removed.

The antibody was conjugated with PerCP-Cy5.5 under optimum conditions, and unconjugated antibody and free PerCP-Cy5.5 were removed. Storage of PerCP-Cy5.5 conjugates in unoptimized diluent is not recommended and may result in loss of signal intensity.

Application Notes

Application

Flow cytometry	Tested During Development
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Recommended Assay Procedure:

A. Stimulation of the Cells

Various *in vitro* methods have been reported for polarization or stimulation of T helper cell subsets of which PMA (Phorbol ester) plus Ionomycin (Calcium Ionophore) has been particularly useful for quickly inducing and characterizing polyclonal cytokine-producing cells. For this kit we recommend the stimulation of mouse spleen cells at a concentration of 1-10 million cells per ml in media for 5 hours with PMA/Ionomycin (at 50 ng/ml and 1 μ g/ml respectively) in the presence of BD GolgiStop™ Protein Transport Inhibitor (provided in the kit or Cat #554724). Add 4 μ l of BD GolgiStop™ for every 6 ml of cell culture and mix thoroughly. It is recommended that BD GolgiStop™ not be kept in cell culture for longer than 12 hours.

Note: Kinetic studies need to be performed to determine the optimal incubation time for each experimental system. Depending on the mouse, frequencies of cytokine producing cells derived from activation of mouse spleen cells can vary widely for a specific cytokine. In particular, the number of IL-17 producing cells can be very low or even negligible on PMA/Ionomycin stimulated spleen cell. In these cases, Th17 polarization cultures should be considered. For specifics on polarization of Th17 cells please refer to the references.

B. Staining of the Cells

1. Harvest of the Cells

Collect cells from *in vitro* stimulatory cultures treated with a protein transport inhibitor. Spin down cells at 250 x g for 10 minutes at room temperature (RT) and wash two times with stain buffer (FBS) (Cat# 554656). Count cells and transfer approximately 1 million cells to each flow test tube (Cat# 352008) for immunofluorescent staining. Cells should be protected from light throughout the staining procedure and storage.

2. Fixing the Cells

- Spin down cells at 250 x g for 10 minutes at RT and thoroughly suspend cells with 1ml of cold BD Cytotfix™ buffer (provided in the kit or Cat# 554655) and incubate for 10-20 minutes at RT.

Note: Cell aggregation can be avoided by vortexing prior to the addition of the fixative.

- Spin down cells at 250 x g for 10 minutes at RT.

Note: After fixation, the cell pellet after centrifugation is loose and care should be taken when aspirating the wash buffer from the tubes. Do not aspirate ALL of the buffer but leave 50-150 μ l of solution in the tubes to avoid cell loss for all subsequent wash steps below.

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- c. Wash cells twice at RT in stain buffer (FBS) and spin down the cells at 250 x g for 10 minutes at RT.

Note: Cells can be stored in stain buffer at 4°C for up to 72 hours or in 90% FCS/10% DMSO at -80°C for up to six months (for samples that need to be stored longer than six months, we recommend performing stability studies).

3. Permeabilizing the Fixed Cells

- a. For cells kept at 4°C, spin down cells at 250 x g for 10 minutes at RT and remove stain buffer.
- b. For cells stored at -80°C thaw and wash twice with stain buffer (FBS) to remove DMSO.
- c. Dilute 10x BD Perm/Wash™ buffer (provided in the kit or Cat# 554723) in distilled water to make a 1x solution prior to use.
- d. Suspend cells in 1ml of 1x BD Perm/Wash™ buffer and incubate at RT for 15 minutes.
- e. Spin down cells at 250 x g for 10 minutes at RT and remove supernatant.

4. Staining with the Cocktail

- a. Thoroughly suspend fixed/permeabilized cells in each tube in 50 µl of BD Perm/Wash™ buffer and add 20 µl/tube of cocktail or appropriate negative control. Incubate at RT for 30 minutes in the dark. Cells should be protected from light throughout the staining procedure.
- b. Optional: Staining of Additional Cell Surface Antigens

Note: If instrument allows, optional multicolor staining of different cell surface antigens can be done at this time. Example: APC-Cy™7 CD3 (Clone 145-2C11, Cat# 557596) and V450 CD44 (CloneIM7, Cat# 560451) etc. For cell surface staining after fixation/permeabilization, suitable antibody clones that recognize denatured epitopes need to be identified.

Note: For antibodies that do not recognize fixed/denatured cell surface markers, it is recommended that staining be done on live cells PRIOR to fixation/permeabilization.

- c. Wash cells twice with 1ml of 1x BD Perm/Wash™ buffer at RT and suspend in Stain Buffer (FBS) prior to flow-cytometric analysis.

C. Flow Cytometric Analysis

Set PMT voltage and compensation using unstained cells and appropriate cell surface markers or use BD™ Compensation beads (Cat #552844) as per the recommended protocol.

Note: It has been reported that CD4 expression on T cells is decreased after cell activation.

Note: Acquire at least 20,000 to 30,000 CD4 positive lymphocytes. Depending on the donor, frequencies of cytokine producing cells derived from activation of human PBMCs can vary widely for a particular cytokine. In order to make statistically significant frequency measurements, sufficiently large sample sizes should be acquired during flow cytometric analysis. Bivariate dot plots or probability contour plots can be generated upon data reanalysis to display the frequencies of and patterns by which individual cells co-express certain levels of cell surface antigen and intracellular cytokine proteins.

Warnings & Precautions

Danger: BD GolgiStop™ Protein Transport Inhibitor (component 51-2092KZ) contains 99.61% ethanol (w/w) and 0.26% monensin, mononatriumsalz (w/w).

Hazard statements:

Highly flammable liquid and vapor.

Causes serious eye irritation.

Precautionary statements:

Keep away from heat/sparks/open flames/hot surfaces. No smoking.

Wear protective gloves / eye protection.

Wear protective clothing.

IF ON SKIN (or hair): Remove / Take off immediately all contaminated clothing. Rinse skin with water / shower.

IF IN EYES: Rinse cautiously with water for several minutes. Remove contact lenses, if present and easy to do. Continue rinsing.

Dispose of contents / container in accordance with local / regional / national / international regulations.

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Danger: BD Cytofix™ Fixation Buffer (component 51-9006613) contains 4.2% (w/w) formaldehyde.

Hazard statements:

Harmful if inhaled.

Causes skin irritation.

Causes serious eye damage.

May cause an allergic skin reaction.

Suspected of causing genetic defects.

May cause cancer. Route of exposure: Inhalative.

May cause respiratory irritation.

Precautionary statements:

Wear protective clothing / eye protection.

Wear protective gloves.

Do not breathe mist/vapours/spray.

IF IN EYES: Rinse cautiously with water for several minutes. Remove contact lenses, if present and easy to do.

Continue rinsing.

If skin irritation or rash occurs: Get medical advice/attention.

Suggested Companion Products

Catalog Number	Name	Size	Clone
554656	Stain Buffer (FBS)	500 mL	(none)
554684	FITC Rat IgG1, κ Isotype Control	0.1 mg	R3-34
554685	PE Rat IgG1, κ Isotype Control	0.1 mg	R3-34
557596	APC-Cy™7 Hamster Anti-Mouse CD3e	0.1 mg	145-2C11
560451	V450 Rat anti-Mouse CD44	0.1 mg	1M7
553066	APC Hamster Anti-Mouse CD3e	0.1 mg	145-2C11
552844	Anti-Rat Ig, κ /Negative Control Compensation Particles Set	6 mL	G16-510E3

Product Notices

1. Please observe the following precautions: Absorption of visible light can significantly alter the energy transfer occurring in any tandem fluorochrome conjugate; therefore, we recommend that special precautions be taken (such as wrapping vials, tubes, or racks in aluminum foil) to prevent exposure of conjugated reagents, including cells stained with those reagents, to room illumination.
2. PerCP-Cy5.5 is optimized for use with a single argon ion laser emitting 488-nm light. Because of the broad absorption spectrum of the tandem fluorochrome, extra care must be taken when using dual-laser cytometers, which may directly excite both PerCP and Cy5.5™. We recommend the use of cross-beam compensation during data acquisition or software compensation during data analysis.
3. PerCP-Cy5.5-labelled antibodies can be used with FITC- and R-PE-labelled reagents in single-laser flow cytometers with no significant spectral overlap of PerCP-Cy5.5, FITC, and R-PE fluorescence.
4. This PerCP-conjugated product is sold under license to the following patent: US Patent No. 4,876,190.
5. Cy is a trademark of Amersham Biosciences Limited. This conjugated product is sold under license to the following patents: US Patent Nos. 5,486,616; 5,569,587; 5,569,766; 5,627,027.
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7. Source of all serum proteins is from USDA inspected abattoirs located in the United States.
8. Caution: Sodium azide yields highly toxic hydrazoic acid under acidic conditions. Dilute azide compounds in running water before discarding to avoid accumulation of potentially explosive deposits in plumbing.
9. For fluorochrome spectra and suitable instrument settings, please refer to our Multicolor Flow Cytometry web page at www.bdbiosciences.com/colors.
10. Please refer to www.bdbiosciences.com/pharming/protocols for technical protocols.

References

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Zhu J, Paul WE. CD4 T cells: fates, functions, and faults. *Blood*. 2008; 112(5):1557-1569. (Biology)

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